

The *rbcL* gene sequence from chestnut indicates a slow rate of evolution in the Fagaceae¹

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The nucleotide sequence was obtained for the chloroplast gene coding for the large subunit of the ribulose 1,5-bisphosphate carboxylase (*rbcL*) of chestnut (*Castanea sativa* Mill.), a member of the woody family Fagaceae. Amplification primers downstream and upstream the *rbcL* open reading frame are also described. By comparing with other angiosperm sequences, we show that the rate of evolution of *rbcL* in the family Fagaceae is much slower than that observed for the families of annuals analyzed.

Key words: angiosperms, *Castanea sativa*, Fagaceae, phylogeny, *rbcL*.

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La séquence nucléotidique du gène chloroplastique codant pour la grosse sous-unité de l'enzyme ribulose 1,5-diphosphate carboxylase (*rbcL*) a été déterminée pour le châtaignier (*Castanea sativa* Mill.), un membre de la famille ligneuse des Fagaceae. Des amores d'amplification en amont et en aval du cadre de lecture de *rbcL* sont aussi décrites. La comparaison avec d'autres séquences d'angiospermes a permis de démontrer que le taux d'évolution du gène *rbcL* dans la famille des Fagaceae est beaucoup plus lent que celui observé chez les familles d'annuelles analysées.

Mots clés : angiospermes, *Castanea sativa*, Fagaceae, phylogénie, *rbcL*.

DNA sequence comparisons of chloroplast-encoded genes is now routinely used to investigate phylogenetic relationships among angiosperms and other plants (Wolfe et al. 1989). In recent years, there has been a growing interest in using sequences of the gene coding for the large subunit of ribulose 1,5-bisphosphate carboxylase/oxygenase (*rbcL*) to estimate plant phylogenies (Ritland and Clegg 1987). The substitution rate of *rbcL* appears appropriate for studies involving taxa that diverged from 10's of million to 100's of million years (Zurawski et al. 1984). In this paper, we report the complete nucleotide sequence of the *rbcL* open reading frame (ORF) for a woody angiosperm, the chestnut of the family Fagaceae, *Castanea sativa* Mill. We show that the rate of evolution of *rbcL* in this family is much slower than that observed for families of annual angiosperms. Implications for phylogenetic studies using *rbcL* are briefly discussed.

rbcL was amplified symmetrically following previously published procedures (Bousquet et al. 1992a). Seven primers, internal to the ORF, were used for amplification and sequencing and were designed from the alignment of previously published ORF sequences of dicots (Bousquet et al. 1992b): for sequencing the 5'-3' strand, 334⁺; 5'-TC TGT TAC TAA CAT GTT TAC TTC-3'; 691⁺; 5'-GA AAC AGG TGA AAT CAA AGG GCA TTA-3'; 1144⁺; 5'-GG TAT TCA CGT TTG GCA TAT GCC TGC-3'; and for sequencing the opposite strand, 216⁻; 5'-TC GGT CCA CAC AGT TGT CCA TGT-3'; 537⁻; 5'-CC CAA

TTT AGG TTT AAT AGT ACA TCC-3'; 979⁻; 5'-AAT ATG ATC TCC ACC AGA CAA ACG TAA-3'; 1303⁻; 5'-TCC CTC ATT ACG AGC TTG TAC ACA-3'. Primers upstream and downstream the coding region were designed from comparing Hamamelideae sequences (Bousquet et al. 1992a): upstream the primer -34⁺ was complementary to the 3'-5' strand, starting at the position -34: 5'-AAG TTC TTA ATT CAT GAG TCG TAG-3'; downstream the primer 1517⁻ was complementary to the 5'-3' strand, starting at the position 1517: 5'-TTA GTA AAA GAT TGG GCC GAG TTT-3'. Direct sequencing of the two DNA strands was performed with the dideoxynucleotide chain-termination procedure using the Sequenase version 2.0 kit with dITP nucleotide mixes (USB) as recommended by the manufacturer, except that times and temperatures were modified (Zimmerman and Fuscoe 1991).

The coding region of the chestnut *rbcL* was 1428 bp long. The chestnut sequence was very similar to red oak (*Quercus rubra*) (Bousquet et al. 1992a) (Fig. 1), another member of the family Fagaceae, with only 12 nucleotide differences for a DNA homology of 99.1%. Otherwise, the chestnut sequence was 90.1% identical to tobacco (*Nicotiana tabacum*) (Shinozaki and Sugiura 1982), 86.5% to maize (*Zea mays*) (Gaut et al. 1992), and 83.5% to Douglas-fir (*Pseudotsuga menziesii*) (Hipkins et al. 1990).

Synonymous and nonsynonymous rates of evolution and their standard errors in different annual and woody families of dicots and monocots were calculated (Li et al. 1985). Overall rates of substitution were estimated using a weighted average of K_s and K_a , the two-parameter method of Kimura (1980), and the one-parameter method

¹The nucleotide sequence reported appears in the GenBank and EMBL under the accession number M94936.

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| | | |
|------|---|------|
| Cs 1 | ATG TCA CCA CAA ACA GAG ACT AAA GCA AGT GTT GGA TTC AAA GCT GGT GTT | 51 |
| Qr 1 | --- | 51 |
| 52 | AAA GAT TAT AAA TTG ACT TAT tAT ACT CCT GAC TAT CAA ACC AAA GAT ACT | 102 |
| 52 | --- | 102 |
| 103 | GAT ATC TTG GCA GCC TTC CGA GTA ACT CCT CAA CCT GGA GTT CCG CCg GAG | 153 |
| 103 | --- | 153 |
| 154 | GAA GCA GGG GCC GCG GTA GCT GAA TCT TCC ACT GGG ACA TGG AgA ACT | 204 |
| 154 | --- | 204 |
| 205 | GTG TGG ACT GAC GGG CTT ACC AGT CTT GAT CGT TAC AAA GGA CGA TGC TAC | 255 |
| 205 | --- | 255 |
| 256 | CAC ATC GAG CCg GTT GCT GGA GAA GAA AAT CAA TTT ATT GCT TAT GTA GCT | 306 |
| 256 | --- | 306 |
| 307 | TAC CCC TTA GAC CTC TTT GAA GAA GGT TCT GTT ACT AAC ATG TTT ACT TCC | 357 |
| 307 | --- | 357 |
| 358 | ATT GTG GGT AAT GTA TTT GGA TTC AAG GCC CTG CGC GCT CTA CGT CTG GAG | 408 |
| 358 | --- | 408 |
| 409 | GAT TTG CGA ATC CCT ACT TCT TAT TCT AAA ACT TTC CAA GGT CCG CCT CAT | 459 |
| 409 | --- | 459 |
| 460 | GGC ATC CAA GTT GAG AGG GAT AAA TTA AAC AAG TAT GGC CGC CCC CTA TTA | 510 |
| 460 | --- | 510 |
| 511 | GGA TGT ACT ATT AAA CCT AAA TTG GGA TTA TCC GCT AAG AAT TAC GGT AGA | 561 |
| 511 | --- | 561 |
| 562 | GCA GTT TAT GAA TGT CTC CGC GGT GGG CTT GAT TTT ACC AAA GAT GAT GAG | 612 |
| 562 | --- | 612 |
| 613 | AAC GTT AAT TCC CAA CCA TTT ATG CGT TGG AGA GAC CGT TTC cTA TTT TGT | 663 |
| 613 | --- | 663 |
| 664 | GCC GAA GCA ATT TAT AAA GCG CAG GCT GAA ACA GGT GAA ATC AAg GGG CAT | 714 |
| 664 | --- | 714 |
| 715 | TAC TTG AAT GCT ACT GCA GGT ACA TGC GAA GAA ATG ATC AAA AGG GCT GTA | 765 |
| 715 | --- | 765 |
| 766 | TTT GCC AGA GAA CTG GGA GTT CCT ATC GTA ATG CAT GAC TAC TTA ACC GGG | 816 |
| 766 | --- | 816 |
| 817 | GGG TTC ACT GCA AAT ACT AGC TTG GCT CAT TAT TGC CGA GAT AAT GGT CTA | 867 |
| 817 | --- | 867 |
| 868 | CTT CTT CAC ATC CAT CGT GCa ATG CAT GCA GTT ATT GAT CGA CAG AAG AAT | 918 |
| 868 | --- | 918 |
| 919 | CAT GGT ATA CAC TTT CGT GTA CTA GCT AAA GCA TTA CGT ATG TCT GGT GGA | 969 |
| 919 | --- | 969 |
| 970 | GAT CAT ATT CAT GCC GGT ACC GTA GTA GGT AAA CTT GAA GGG GAA AGA GAA | 1020 |
| 970 | --- | 1020 |
| 1021 | ATC ACT TTA GGC TTT GTT GAT TTA CTA CGT GAT GAT TAT ATT GAA AAA GAT | 1071 |
| 1021 | --- | 1071 |
| 1072 | CGA AGC CGC GGT ATT TAT TTC ACT CAA GAT TGG GTC TCT TTA CCA GGT GTT | 1122 |
| 1072 | --- | 1122 |
| 1123 | CTG CCC GTG GCT TCA GGG GGT ATT CAC GTT TGG CAT ATG CCT GCT TTG ACC | 1173 |
| 1123 | --- | 1173 |
| 1174 | GAA ATC TTT GGA GAT GAT TCC GTA CTA CAA TTt GGC GGA GGA ACT TTg GGG | 1224 |
| 1174 | --- | 1224 |
| 1225 | CAC CCT TGG GGA AAT GCA CCT GGT GCc GTA GCT AAT CGA GTA GCT CTA GAA | 1275 |
| 1225 | --- | 1275 |
| 1276 | GCA TGT GTA CAA GCT CGT AAT GAG GGA CGT GAT CTT GCT CGT GAG GGT AAT | 1326 |
| 1276 | --- | 1326 |
| 1327 | GAA ATT ATT CGT GAG GCT GCT AAA TGG AGT CCT GAG CTA GCT GCT TGT | 1377 |
| 1327 | --- | 1377 |
| 1378 | GAA GTA TGG AAA GAG ATC AAA TTT GAA TTC CCA GCA ATG GAT ACT TTG TAA | 1428 |
| 1378 | --- | 1428 |

FIG. 1. Open reading frame sequence (1428 bp) of the *rbcL* gene from *Castanea sativa* Mill. Differences are indicated by lowercase letters between *C. sativa* (Cs) and *Quercus rubra* (Qr) (Bousquet et al. 1992a), another member of the family Fagaceae.

of Jukes and Cantor (1969). For each family analyzed, a rate per year was obtained by dividing the pairwise rate between two taxa by $2T$, where T was the divergence time derived from the fossil record.

Substitution rates per year varied in a range of near one order of magnitude between the two woody and the

two annual angiosperm families compared (Table 1). This difference was apparent at both synonymous and non-synonymous sites (Table 1). The slower rate of evolution observed in the two woody families analyzed confirms a rate heterogeneity trend recently reported among seed plants for *rbcL*, although no rates per year were esti-

TABLE 1. Substitution rates per site per year (and substitution rates per site in parentheses) of *rbcL* within four angiosperm families

| Families | Taxa compared | Divergence time (million years)* | Number of sites compared | Substitution rate \pm SE† | | |
|------------|--------------------------|----------------------------------|--------------------------|--|--|--|
| | | | | K_s | K_a | K_o |
| Fagaceae | <i>Castanea-Quercus</i> | 60 | 1428 | $2.36 \pm 0.79 \times 10^{-10}$ (0.0283 ± 0.0095) | $0.23 \pm 0.13 \times 10^{-10}$ (0.0027 ± 0.0016) | $0.71 \pm 0.21 \times 10^{-10}$ (0.0085 ± 0.0025) |
| Solanaceae | <i>Nicotiana-Petunia</i> | 50 | 1434 | $7.99 \pm 1.64 \times 10^{-10}$ (0.0799 ± 0.0164) | $1.21 \pm 0.33 \times 10^{-10}$ (0.0121 ± 0.0033) | $2.75 \pm 0.45 \times 10^{-10}$ (0.0275 ± 0.0045) |
| Poaceae | <i>Zea-Triticum</i> | 60 | 1428 | $1.34 \pm 2.06 \times 10^{-10}$ (0.1606 ± 0.0247) | $1.90 \pm 0.38 \times 10^{-10}$ (0.0228 ± 0.0046) | $4.45 \pm 0.54 \times 10^{-10}$ (0.0534 ± 0.0065) |
| Arecaceae | <i>Serenoa-Calamus</i> | 60 | 985 | $1.98 \pm 0.89 \times 10^{-10}$ (0.0237 ± 0.0107) | $0.54 \pm 0.24 \times 10^{-10}$ (0.0065 ± 0.0029) | $0.86 \pm 0.28 \times 10^{-10}$ (0.0103 ± 0.0033) |

*References for divergence times: Fagaceae, Crepet (1989); Solanaceae, Albert et al. (1992); Poaceae, Wolfe et al. (1989); Arecaceae, Wilson et al. (1990).

†Abbreviations used: K_s , K_a , and K_o synonymous, nonsynonymous, and overall numbers of substitutions, respectively (Li et al. 1985); Kimura's (1980) two-parameter numbers of substitutions; J.C., Jukes and Cantor's (1969) one-parameter numbers of substitutions.

mated (Bousquet et al. 1992b). This diversity of rates implies that careful analysis of rate heterogeneity among the sequences analyzed must be conducted before using this gene as a molecular clock, which assumes constancy of evolutionary rate over time (Zuckerkandl and Pauling 1965). Procedures for this purpose have recently become available (Wu and Li 1985; Li and Bousquet 1992). Methods of phylogenetic tree construction, which are sensitive to rate heterogeneity (see Sourdis and Nei 1988), must also be used cautiously with this gene.

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