REVIEW SUMMARY

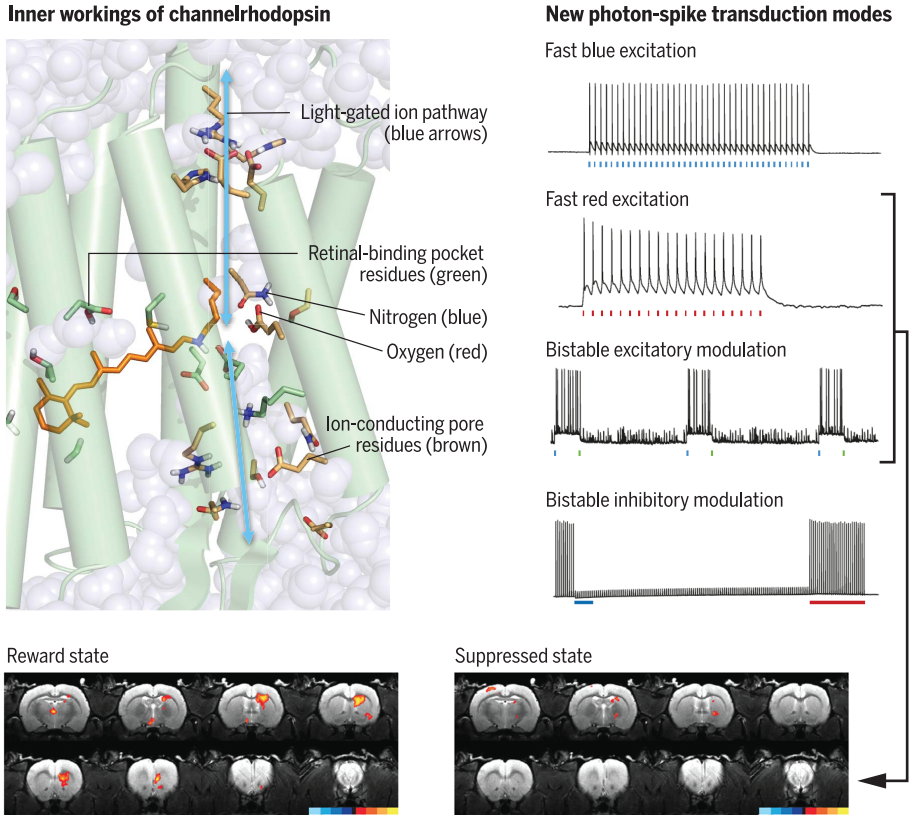
OPTOGENETICS

The form and function of channelrhodopsin

Karl Deisseroth* and Peter Hegemann*

BACKGROUND: Channelrhodopsins (ChRs) are naturally occurring light-gated ion channels that are important for allowing motile algal cells to find suitable light levels. In neuroscience, ChRs have become broadly significant for helping to enable the control of specific circuit elements with light (i.e., optogenetics). Research into how sensation, cognition, and behavior arise from neuronal activity dynamics has been enabled by the expression of ChRs, and other members of the microbial opsin family, in specific cells or in specific connections within nervous systems of behaving animals.

Both the unique light-gated channels themselves and opportunities for their biological application have been under intense investigation. The resulting studies of atomic-scale structure-function relationships have led not only to sophisticated understanding of the underlying chemical processes governing these unique seven-transmembrane channels from the plant kingdom, but also (via optogenetics) to the discovery of fundamental neural circuit principles underlying adaptive and maladaptive behavior in animals.



A light-gated ion pore. (Top) Left: Inner workings of channelrhodopsin. Right: New photon-spike transduction modes arising from structure-guided redesign. (Bottom) Discovering the causal underpinnings of depression-related symptomatology. Brain region–specific activity dynamics of the mammalian dopamine neuron–driven reward state (left) are suppressed by the prefrontal cortex (right) as shown, using the second and third photon-spike transduction modes.

ADVANCES: The atomic-scale understanding of light-gated ion channel function has spanned the key processes of activation/deactivation gating, light adaptation, color tuning, and ion selectivity. A ChR crystal structure–derived, molecular dynamics–calculated pore snapshot (top left panel of the figure) summarizes the wide scope of biophysical and biochemical discoveries. Molecular modeling and redesign have created multiple modes of coupling between delivered photons and spikes in an approach that has illuminated basic principles of protein function and also created new tools for optogenetics. In the top right panel of the figure, the top trace shows a photon-spike transduction mode arising from the ChETA mutation, which results in high-speed, high-fidelity single blue flash–single spike coupling. The second trace shows red photon-spike transduction arising from a redshifted ChR found in nature and then engineered for stronger, more redshifted performance (C1V1). The third trace shows bistable excitation photon-spike logic, in which step-function opsin (SFO) mutations were introduced to create stalled photocycles, allowing stable excitation without continuous light delivery. The bottom trace shows bistable inhibition photon-spike logic; ChRs that are normally cation-conducting, and are therefore excitatory in neural systems, were converted to anion-conducting (inhibitory) ChRs by replacing negatively charged pore residues, followed by SFO mutations for bistability. The C1V1 and SFO designs together allowed us to determine that the medial prefrontal neocortex modulates interactions between two distant subcortical structures to control reward-mediating physiology and behavior (clarityresourcecenter.org/ofMRI.html; www.optogenetics.org).

ON OUR WEBSITE

Read the full article at http://dx.doi.org/10.1126/science.aan5544

OUTLOOK: The ChR light-gated pore will continue to be studied for its own elegant properties, which are paradigmatic among ion channels because light-gated systems allow structure-function analysis on the femtosecond time scale. Meanwhile, psychiatry has already yielded some of its deepest mysteries to ChR pore structural insights, including in explorations of clinically relevant behavioral states such as anhedonia. Many more opportunities for ChRs in basic neuroscience remain untapped, with the potential for precision redesign to achieve new applications and new roles integrated with other advanced technologies. ■

The list of author affiliations is available in the full article online.
*Corresponding author. Email: deissero@stanford.edu (K.D.); hegemann@rz.hu-berlin.de (P.H.)
Cite this article as K. Deisseroth, P. Hegemann, *Science* **357**, eaan5544 (2017). DOI: 10.1126/science.aan5544

REVIEW

OPTOGENETICS

The form and function of channelrhodopsin

Karl Deisseroth[1,2,3]* and Peter Hegemann[4,5]*

Channelrhodopsins are light-gated ion channels that, via regulation of flagellar function, enable single-celled motile algae to seek ambient light conditions suitable for photosynthesis and survival. These plant behavioral responses were initially investigated more than 150 years ago. Recently, major principles of function for light-gated ion channels have been elucidated by creating channelrhodopsins with kinetics that are accelerated or slowed over orders of magnitude, by discovering and designing channelrhodopsins with altered spectral properties, by solving the high-resolution channelrhodopsin crystal structure, and by structural model–guided redesign of channelrhodopsins for altered ion selectivity. Each of these discoveries not only revealed basic principles governing the operation of light-gated ion channels, but also enabled the creation of new proteins for illuminating, via optogenetics, the fundamentals of brain function.

The study of neural circuitry underlying adaptive and maladaptive animal behavior has become intertwined with the investigation of algae-derived light-gated ion channels (*1*, *2*). Each of these two fields has unexpectedly, profoundly advanced the other, culminating in mutually illuminating discoveries (*3*). Curiosity regarding the nature and operation of these unique channels, combined with a long-standing impetus to bring precise cellular-resolution causality to basic and clinical neuroscience, together have resulted in the elucidation both of diverse fundamental processes in neuroscience [reviewed in (*3*)] and of underlying structural and functional mechanisms within the algae proteins themselves (explored here).

The natural role of channelrhodopsins (ChRs) in motile algae involves coupling irradiance information from ambient light to flagellar motion, allowing the organism to seek light optimal for photosynthesis and survival. ChR signaling must be fast enough to implement meaningful coupling to the flagella during helical swimming of the alga with its typical 2-Hz rotation, which continuously alters eye position relative to direction of incident light (*4*). Here, we explore how studies over the past 10 years have elucidated the atomic-scale protein structural underpinnings of this single polypeptide–mediated fast transduction of visible-wavelength photons into selective transmembrane ion flow. We describe how, within these compact proteins, even single amino acids play multiple roles in light-gated ion channel function, including key processes such as activation/deactivation gating, light adaptation, color specificity, and ion selectivity (Fig. 1). In doing so, we bring together structural and functional insights, consider general principles of selective ion channel design and evolution (subject to biophysical constraints that the light-activated ion-conducting pore satisfies), and highlight how the basic discovery process of designing and creating new classes of light-gated ion channels not only provided fundamental insight into channelrhodopsin pore function, but also enabled the discovery of principles underlying nervous system function and neuropsychiatric disease.

[1]Department of Bioengineering, Stanford University, Stanford, CA, USA. [2]Department of Psychiatry and Behavioral Sciences, Stanford University, Stanford, CA, USA. [3]Howard Hughes Medical Institute, Stanford University, Stanford, CA, USA. [4]Institute for Biology, Humboldt Universität zu Berlin, D-10115 Berlin, Germany. [5]Experimental Biophysics, Humboldt Universität zu Berlin, D-10115 Berlin, Germany.
*Corresponding author. Email: deissero@stanford.edu (K.D.); hegemann@rz.hu-berlin.de (P.H.)

Deep roots in botany

The channelrhodopsin story begins at the Neva River in Russia nearly 150 years ago, where the famed 19th-century botanist Andrei Sergeyevich Faminzin studied motile microalgae (Fig. 2, A and B). Faminzin provided the first comprehensive descriptions of single-celled motile algae moving toward or away from light (*5*). Despite intensive study over the next century of behavioral ecology among algal species, and of subcellular light detection structures that algae use to modulate flagellar beating (Fig. 2C, *Chlamydomonas reinhardtii*), the photoreceptor and molecular phototransduction mechanisms remained mysterious. Not until the late 20th century were light-evoked rhodopsin-type currents in *Chlamydomonas*-type chlorophyceae (*6*, *7*), and the broader family of microbial rhodopsin-type ion conductance regulators, detected (*1*, *8*, *9*).

In 1971, evidence for microbial retinal-binding membrane proteins was obtained, initially the proton pumping–type bacteriorhodopsin within its native archaeal system (*Halobacterium salinarum*) (*8*). Purified bacteriorhodopsin-mediated light-activated transmembrane pump currents were later demonstrated in artificial black lipid membrane preparations (*9*). More than 40 years of study revealed that rhodopsins produced by microbial organisms include several subclasses of single gene–single protein ion transporters. All are seven-transmembrane (7TM) retinal-binding proteins encoded by single opsin genes (*1*, *2*). Unlike rhodopsins of the vertebrate retina, these microbial proteins bind all-trans retinal (ATR, rather than cis isomers) in the dark/inactive state (Fig. 3). Rather than triggering an effector cascade to elicit ion flux (like vertebrate rhodopsins), light sensation and ion flow regulation are implemented by a single polypeptide chain (*1*, *2*).

The photocurrents of motile Chlorophyceae are mediated by the ChR subtype of this family; the prototype ChRs are ChR1 and ChR2 of *Chlamydomonas*. These algal rhodopsins were discovered via action spectroscopy of phototaxis/photoelectric responses and via restoration of photobehavior in blind mutant algae by supplementation with retinoids (*7*, *10*). Characterization of algal photocurrents led to the suggestion that light-sensing and ion-conducting units derived from a single protein (*11*) conducting 10 to 100 charges per photocycle [initial 300-fS estimation assuming 10,000 channels in the algal eyespot (*7*) was quite consistent with 100-fS unitary conductances inferred by noise analysis years later for heterologously expressed ChR2]. After photoreceptor-like sequences were discovered in the Kazusa cDNA database, heterologous expression demonstrated passive transmembrane currents down electrochemical gradients (revealing channel-like behavior) (*12*, *13*).

ChRs are larger than any previously identified rhodopsin, but only the N-terminal 40% [including all transmembrane domains with both light sensor and ion channel functionality (*12*)] is needed for optogenetics (*14*). As confirmed by crystal structure determination (*15*), the ATR chromophore appears to have inherited positioning within the retinal binding pocket (RBP) and covalent binding to the protein backbone (via lysine; Fig. 1) of its presumed rhodopsin-pump evolutionary forebears (*1*, *2*).

Studies of motile photosynthetic microbes have continued along several directions. Not all such organisms use ChRs; light-activated nucleotide cyclases with flavin-based light sensors have been found in phototactic Euglenophyceae (*16*). In *Chlamydomonas*, structural and behavioral work is ongoing; ChRs have been localized to the eyespot (photosensitive organelle) overlaying part of the plasma membrane (*17*–*19*) (Fig. 2C), and light modulation of flagellar beating has recently been recorded with unprecedented precision (*20*) as it occurs via flagellar action potentials (upon large and abrupt changes in light intensity) or by gradual modulation of membrane voltage. As detailed next, studies of these behaving plants have also profoundly advanced the understanding of behaving animals (*3*, *14*).

Impetus from neuroscience and basic principles of optogenetics

Francis Crick was the first to suggest that light might be a useful cell type–targeted neural control

modality for the investigation of nervous system function, but did not see how to achieve this, terming the possibility "far-fetched" (*21*). Over the decade beginning in 2005, light-activated proton or Cl^- pumps (bacteriorhodopsins or halorhodopsins) and light-activated cation or Cl^- channels (ChRs) were all discovered to serve as high-speed optically activated regulators of action potential firing when genetically targeted to neurons of metazoa (*14*). Together with several additional (and necessary) technological innovations, this enabled precisely what Crick had sought (*3*, *14*).

Targeted optical control of single neurons had been achieved earlier with spatially guided lasers but not broadly applied (*22*, *23*). Early genetically guided efforts were elegant but limited as well, in part because their multicomponent design required multiple genes, or both exogenous chemicals and effector genes, to couple the chromophore to neuronal activity (*24*–*26*). Reports of the feasibility of a single-component microbial opsin approach to optical control of spiking were initially published only in transparent or spatially superficial systems (e.g., cultured neurons, isolated retina preparations, nervous system slices, and small invertebrates) (*27*–*31*). Justified skepticism remained for many years regarding the potential for general utility across intact neural systems.

These microbial opsin genes constitute only one feature of what became the optogenetic approach. Two other key developments—the fiber-optic neural interface and versatile targeting methodology (*32*–*37*)—would ultimately enable generalizable millisecond-precision genetically targeted neuronal control with light throughout the brain during behavior (*3*, *14*). By 2007, optogenetic control of defined spiking patterns in specified neurons deep in the hypothalamus of freely moving adult mice had been achieved, along with resulting behavioral state transitions (*33*). Optogenetics had attained an early form of the fundamental methodology that neuroscientists use today: expression of microbial opsins with cell type (and even projection type) specificity and behavioral control potency by means of high-titer viral gene-targeting vectors and light targeting through implanted fiber optics (*34*). By 2009, generalizable gene-targeting strategies had been developed and shown to be suitable for mammalian behavioral control (*35*). From across the global scientific community, many thousands of discoveries have resulted, revealing the otherwise inaccessible cell-specific neural activity building blocks of behavior (*3*, *14*).

Structural models and pore-gating kinetics

The light-activated membrane pore of ChRs is unique. Within the ChR family, however, many variants exist. ChR is a highly flexible protein heavily modified in action spectrum, photocycle kinetics, and ion selectivity by scientists over the past dozen years, but much more so by nature on the billion-year time scale.

There was initial confusion regarding the structure of the light-activated pore. Early characterization of ChRs had elucidated properties such as nonspecific cation selectivity (permeability to $H^+ >> Na^+ > K^+ >> Ca^{2+}$ with inward rectification) (*12*, *13*). But it was unclear where the pore would be localized, except for perhaps a contribution from helix 2, with its prominent intramembranous glutamate residues. In the quest to find the light-gated pore, comparisons with microbial light-driven pump structures were of little help—for example, helices 1 and 2 show very low homology with presumptive pump counterparts—or even misleading [low-resolution cryo–electron microscopy (cryo-EM) had led to predictions that the pore would reside at the interface of two ChR monomers (*36*)—which turned out to not be the case (*15*)]. This level of uncertainty stalled pore engineering for design and creation of novel ion-selective ChRs. Many other functions, including the key kinetic properties of activation and inactivation (Fig. 3), nonetheless yielded to construction of structural models and detailed molecular design before the three-dimensional structure was solved.

Gradual inactivation of ChR photocurrents in strong light (Fig. 3) appears useful for algae in coping with high light intensities but poses challenges for neuroscience. This inactivation manifests as a probabilistic alternative photoisomerization around the ATR C15=NH bond (*37*), corresponding

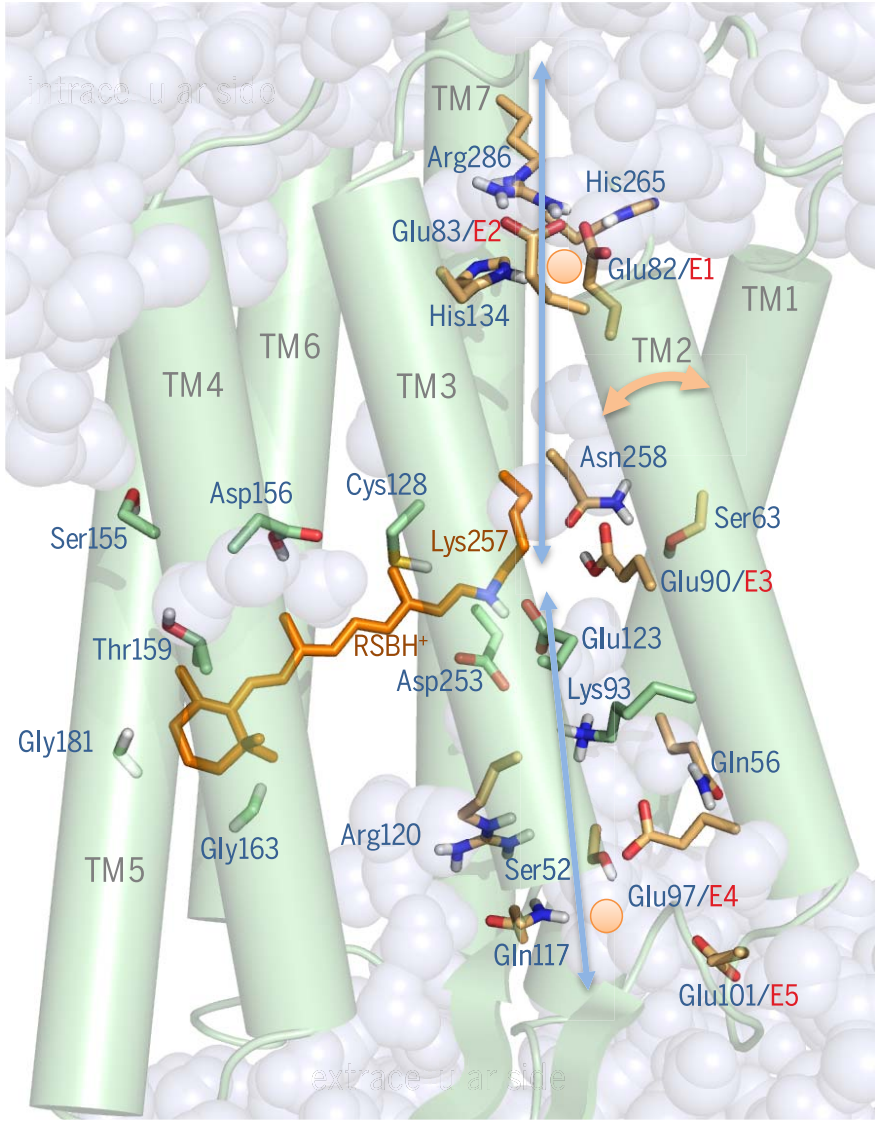


Fig. 1. The light-gated pore. The crystal structure of C1C2 is shown as a 3UG9[13]-derived pore snapshot calculated via molecular dynamics (MD). Protonated retinal Schiff base (RSBH+, orange) and key pore (brown) or RBP (green) residues showing protonation states of polar residues (*81*) are shown in stick form (red, oxygen; blue, nitrogen; white, polar protons). ChR2 residue numbering is used. Displayed waters invaded from both membranes during calculations (*81*). Two Na^+ ions (orange circles) are shown at positions of preferred occupation within the access channel and near inner gate calculated by MD (*81*). Blue arrows denote the presumptive permeation pathway. The two-headed orange arrow shows the predicted (*43*) tilting dynamic of helix 2. Conserved glutamates in helix 2 are designated E1 to E5.

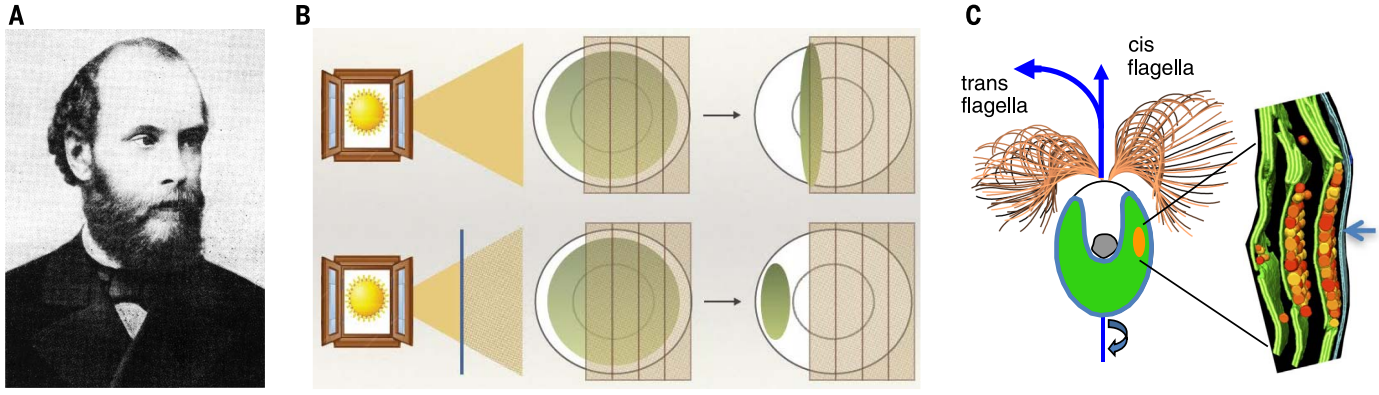


Fig. 2. Deep roots in botany. (**A**) Faminzin, pioneer of algal behavior (*4*). (**B**) Faminzin's landmark observation (*5*): At low light intensities (bottom), *Chlamydomonas* exhibits phototaxis toward light, whereas at higher light intensities (top) the algal cells accumulate at optimal irradiance. (**C**) Adapted cartoons showing subcellular *Chlamydomonas* structure: flagellar beating pattern (*20*) and eye structure reconstructed from high-resolution tomography (*17*). Eyespot overlaying part of plasma membrane (arrow) corresponds to ChR location. Layers of carotenoid-containing vesicles (colored spheres) held by chloroplast membranes (green) serve as an optical device (interference reflector) (*4*, *19*).

to transition from a syn-cycle to an anti-cycle (Fig. 3) as in animal rhodopsins during transitions from Meta-I to Meta-III (*38*) or in bacteriorhodopsin during dark adaptation (*39*). The syn-cycle open state is less permeable to protons and cations (versus the anti-cycle), contributing to photocurrent decline under prolonged illumination (Fig. 3) (*40*). Under dark adaptation, the ATR exists as all-trans/15-anti, but shortly after illumination a mixture of two similar but distinguishable states appears (all-trans/15-anti and 13-cis/15-syn), leading to conducting states in 13-cis/15-anti (O1) and 13-trans/15-syn (O2) configurations, respectively (*37*, *41*). The O1/O2 ratio also depends on membrane voltage and pH gradient, and differs substantially among ChR variants, contributing to diversity of photocurrent properties (*1*, *2*, *42*).

The fast time constant for actuation of pump rhodopsins known from earlier work was concordant with later-observed ChR activation in oocyte membrane patches (τ_{on} < 250 μs) (*13*), and elucidation of molecular principles governing pore kinetics became a major goal, not only to obtain insight into the mechanics of pore operation but also to achieve optogenetic control over both short and long time scales of behavior and physiology. Retinal isomerization appears to cause rapid rearrangement of central gate residues and gate opening on the nanosecond time scale; helix hydration via water influx proceeds within 10 to 100 μs, which (in conjunction with helix 2 movement) opens an inner gate, allowing formation of the cation-conducting pore (*43*, *44*) (Fig. 1). Under typical conditions, the submillisecond opening time constant (τ_{on}) of the ChR pore is fast enough not to influence or limit neuroscience applications. In contrast, photocurrent decay after light-off (τ_{off}) is of great functional consequence in neuroscience and is broadly tunable. Pumps and channels will cease to enter the photocycle upon cessation of light, whereas observed photocurrents can outlast light because already activated proteins are completing their photocycle. The τ_{off} for wild-type ChR2 (~10 ms) is impressively fast for a plant, but is slow for fast-spiking mammalian neurons and contributes to impaired fidelity in optogenetics (*45*). This parameter [which in many ChRs further slows upon membrane depolarization (*42*, *46*)] elicits prolonged post-spike depolarization lasting tens of milliseconds, counteracting typical millisecond-scale repolarizations that terminate natural spikes; artifactual doublets rather than precise single spikes can result (*42*, *45*), and spiking can even fail within higher-frequency spike trains (because repolarization de-inactivates native voltage-dependent channels required for subsequent spiking) (*42*, *45*). These fidelity issues derived from long ChR τ_{off} values (*45*) thus were addressable with redesigned shorter τ_{off} once key molecular principles had been determined.

Mutations at Glu^{123} of ChR2, one of the negatively charged "counterion" residues stabilizing the obligate intramembranous positive charge of the protonated retinal Schiff base ($RSBH^+$; Fig. 1 and Fig. 4, A and B), reduce τ_{off} to 4 ms at −100 mV and also reduce voltage-dependent slowing, thus accelerating pore closure during spiking (*45*). These "ChETA" variants (*45*) exhibited the drawback of moderately reduced light sensitivity due to reduced photoisomerization efficiency and charge transfer per photon (*45*). However, accelerated deactivation addressed the key parameter limiting neuroscience application to fast-spiking cells and enabled firing at 200 Hz or more, along with reduced numbers of extra or missed spikes (*42*, *45*) (Fig. 4B). These high-speed variants are often used when precise control and temporal stationarity is desired, particularly in fast-spiking cells [involved in sleep, fear, and feeding (*46*–*49*)]. Further engineered or naturally occurring ChRs were later identified that also exhibited fast deactivation and/or improved temporal stationarity [e.g., ChIEF, Chronos, and other variants; reviewed here (*14*, *41*, *42*, *50*, *51*)], and the fast-ChR toolkit grew further as ChETA and ChIEF modifications proved portable to other ChRs, including designer variants with larger photocurrents (*51*) and redshifted spectra (discussed below).

Bistable modes of pore operation

What about extending rather than shortening photocycles? In a surprising discovery, slowing τ_{off} was even more useful than accelerating τ_{off} in achieving kinetic pore bistability (*52*). These modified ChRs did not require continuous light for continuous function, but instead were bistable as a result of precisely reversible photocycle arrest (Fig. 4F) (*52*). Similarly arresting pump photocycles would mean termination of photocurrent, but for channels, precision kinetic locking into an open-pore state enables stable current for tens of minutes after light-off (*52*). Modification of helix 3–helix 4 interaction at the DC pair (Cys^{128}-Asp^{156} in ChR2; Fig. 4, E and F) extends open-state lifetimes by up to 6 log units and reduces inactivation (*52*–*54*). Interestingly, redshifted light excitation of the resulting stable conducting state reconverts ATR back to the dark-state conformation, causing channel closure (*52*). Thus, high temporal precision for both onset and offset is preserved, with intervening step-like performance stable enough to make continuous light unnecessary (Fig. 4F); these variants were thus termed step-function opsins (SFOs) (*52*).

Several advantages accrue from application of SFOs to optogenetics: (i) Light need not be provided continuously. The many-orders-of-magnitude reduction (*42*, *52*) in energy delivered can eliminate any intense-light toxicity from long-term experiments. (ii) If complex behaviors (e.g., large-group social interactions) or experimental settings [e.g., naturalistic environments or magnetic resonance imaging (MRI) scanners] are incompatible with certain devices, cells can be stably modulated with a light pulse, followed by characterization of physiology or behavior over prolonged periods without light delivery hardware (*54*). (iii) Continuous mild depolarization can simply favor excitability of targeted cells [rather than provide precise user-defined spikes (*52*, *54*)], an approach often leveraged to maintain or enhance native rhythm/timing relationships (*54*–*61*). (iv) Cells expressing these bistable ChRs exhibit vastly

greater light sensitivity (42, 52–54) due to photon integration; physiologically relevant excitatory photocurrents of >100 pA can be elicited with wild-type ChRs at safe expression levels using ~1 mW/mm^2 irradiance at the expressing neurons, but cells expressing the SFO ChR2-C128A/S variants (τ_{off} up to three orders of magnitude longer) are 300 times as light-sensitive (52) for stationary photocurrents. Because bistable channels are slow to close, photocurrent accumulates even with orders-of-magnitude weaker light. Even further reduced energy delivery (over time and instantaneously), as well as greater tissue volumes recruited at a given light intensity in large-brained subjects, can result from the use of DC double mutations [the most stable SSFO (stabilized SFO) variant (54) with $\tau_{off} \geq 30$ min; Fig. 5A]. This phenomenon has been leveraged to minimize heating (and heating artifacts) during optogenetic functional MRI (fMRI) (Fig. 5B) (56).

Numerous applications leverage these SFO capabilities (54–61). Indeed, SFO mutations, like the ChETA/CHIEF mutations described above, are portable to certain other ChR backbones for new functionality. For example, combined with His^{134} → Arg (13), a mutation that increases photocurrents [as do Thr^{159} → Cys (51), Thr^{59} → Ser (62), and Thr^{246} → Asn (62)], the SFO strategy creates two-photon illumination–recruited bistable ChRs (2PSFOs) (63). Furthermore, when combined with Cl^--conducting ChRs as discussed below, bistable inhibition results (Fig. 4G) (62, 64), with versatile utility [e.g., in studying pain circuitry (60)].

Spectral properties

Fast, efficient structural changes require tight protein-chromophore contact and thus RBP structural rigidity; such fast changes are important not only for kinetics, but also for spectral properties of ChR pore gating. Absorption peaks for known ChRs span 440 to 590 nm (1, 41, 50, 65), and 630- to 644-nm peaks are seen in certain other retinal-binding proteins (66, 67). Absorption spectra are determined by factors including RBP polarity, ATR planarity [in particular, coplanarity of the β-ionone ring C6=C7 bond with the polyene chain in the 6-s conformation (68)], and connection of negatively charged RSBH counterions with long-range hydrogen-bonding networks (68–70) (Figs. 1 and 4).

ChR spectra continue to drive basic and applied investigation. Certain bands of lower-energy (red) light can penetrate somewhat more deeply and safely into biological tissue than blue light. Combinatorial control is also enabled using two-color strategies (54); moreover, all-optical activity-guided/closed-loop experiments become more straightforward when redshifted control tools can be integrated with robust blue light–activated optical readouts (3, 50, 71). But the extent to which RBP-level understanding from other redshifted rhodopsins (69) could drive insights into ChR itself, or into development of color-shifted optogenetic tools, was initially unclear. Despite the recent emergence of additional RBP structural information from a color-shifted ChR (68), attempts over years in many laboratories to build and leverage structural models to engineer or evolve usefully redshifted (by ~100 nm) ChRs were unsuccessful, perhaps because of the predominance of multifactorial and long-range contributions to spectral properties evolved over hundreds of millions of years.

The first discovery (70) of a substantially redshifted ChR, VChR1, finally came in the alga *Volvox carteri* (via searching for opsins in U.S. Joint Genome Institute databases). Initial characterization in culture (70) revealed that a previously inaccessible window was now available, >560 nm, where *Chlamydomonas* ChRs (peaking at ~470 to 490 nm) exhibit almost zero photocurrent. Universally, ChRs (including VChR1 and all other ChRs identified or engineered since) retain robust responsivity in a short-wavelength/blue shoulder of the action spectrum (50) (~470 nm). Persistent blue actuation appears to be an intrinsic property of the retinal polyene system [relating to activation of higher electronic state transitions beyond the lowest-energy transition available (71)]. Although VChR1 photocurrents were small (<100 pA) (70), several modifications in combination (Fig. 1)—including provision of membrane-trafficking/endoplasmic reticulum export motifs identified earlier for enabling halorhodopsin optogenetics (72, 73), chimerization with ChR1 elements (74), and in some cases mutations reducing the blue shoulder and/or ChETA mutation (45)—resulted in diverse members of the new C1V1 ChR family (54). In 2011, C1V1 enabled the first red light–driven spiking (Fig. 4D) (54) as well as in vivo combinatorial optogenetics [two populations separably controlled with red and blue light (54)], which allowed the long-sought demonstration in mouse experiments a causal role for excitation-inhibition balance in governing gamma oscillations and social behavior (54).

Another application emerged with (i) the discovery of high responsivity of red light–driven opsins to two-photon illumination, enabling single-cell resolution optogenetics in brain tissue (63), and (ii) integration of red light–excited control with blue light–excited readout (via genetically encoded activity sensors such as GCaMP Ca^{2+} reporters). Enabling these in vivo all-optical play-in/read-out experiments (50, 75–78) has opened the door to tuning optogenetic control in order to match timing and amplitude of naturally occurring activity in the same circuit elements (3, 76), and more broadly to keeping stimulation attuned to native dynamics and events in real time through closed-loop and activity-guided strategies (50, 74–78). Many opsins are now available for redshifted excitation, including not just VChR1 and C1V1 but also the VChR1-based ReaChR (79) as well as MChR1 (80), Chrimson (65), and bReaCHES (77) (with ChETA modifications for speed). As with earlier insights into kinetics, the initial VChR backbones and C1V1 modifications provided not only insight into ChR structure-function relationships (Fig. 4), but also new neuroscience functionality [for example, enabling discovery of the causal role of the medial prefrontal cortex (mPFC) in regulating midbrain–nucleus accumbens interactions and reward-related behavior (56) (Fig. 5)].

High-resolution crystal structure

Although the above innovations were enabled by ChR modeling without complete structural knowledge, major domains of discovery and design remained difficult to address without high-resolution information. After many years and the creation of numerous constructs for expression and crystallization, in 2012 the 2.3 Å crystal structure was obtained for a truncated functional chimera between *Chlamydomonas* ChR1 and ChR2 (C1C2)

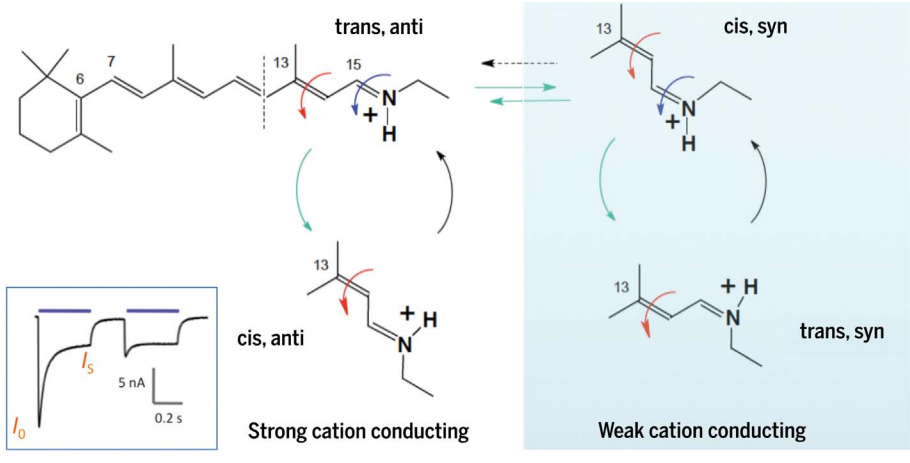


Fig. 3. Chromophore states and channel currents. Chromophore configurations in RSB: all-trans, 15-anti (trans, anti) of dark-adapted state; 13-cis, 15-syn (cis, syn) of second dark state (increasingly occupied after blue illumination during photocurrent inactivation from initial I_0 to stationary I_S; inset) (1, 37). In corresponding open states occupied after $^{13}C{=}^{14}C$ photoisomerization, RSB is in the 13-cis, 15-anti (cis, syn) configuration in the open conducting state O1 and 13-trans, 15-syn (trans, syn) in O2 (1, 37). Cis-trans isomerization is indicated by red arrows, anti-syn isomerization by blue arrows, photochemical conversions by green arrows, and thermal conversions by black arrows.

(15). This structure revealed many of the deepest mysteries of the channel, including its long-sought internal light-activated pore (Fig. 1); as anticipated, questions were also raised and new ideas emerged.

Although the structure revealed a covalently linked dimer of 7TM proteins (associated by N termini and helices 3 and 4) (15), no pore was present at the interface (15), contrary to prior prediction (36). Rather, monomers displayed internal pores formed from four of the seven transmembrane helices (TMs 1, 2, 3, and 7; Fig. 1), with two tilted to create space for a partially water-filled polar channel with two closed-state pore gates requiring repositioning for creation of ion-conducting states. Close to the intracellular membrane, the inner gate is formed by two highly conserved glutamates (E1 and E2; Fig. 1), which interact with His^{134} and His^{265}, closing the channel to both water and ions in the dark. Interestingly, this site is also a key determinant of cation/proton selectivity (severely impaired by E1/E2 or His^{134} mutation). The central gate [structurally separated from the inner by a water-deprived vestibule, as also supported by QM/MM (quantum mechanics/molecular mechanics) calculations (81) (Fig. 1)] consists of Ser^{63}, Asn^{258}, and E3 of Fig. 1, locking the protein from the extracellular bulk phase with respect to water/proton/cation flux and serving as a second selectivity filter even though separated from the surface (15) by 8 Å.

Other key insights included revelation of the RBP and exact side chain positioning defining the chromophore electrostatic environment (15). RBP structure has numerous implications for understanding the channel, including gating regulation between conducting and nonconducting states, ion selectivity, and spectral properties. ChR photocurrents exhibit a transient peak (I_p) decaying to stationary (I_s) under continuous illumination (Fig. 3); for conducting states (O1, O2), most channels reside in O1 during I_p, but under prolonged stimulation, low-conduction O2 accumulates and photocurrent decreases. These transitions likely involve protonations and partial charge-localization shifts among chromophore-proximal residues, overlapping with potential targets for shifting spectral properties.

In natural and artificial ChRs with bathochromic (red-shifted) absorption, the RBP is made more polar near the RSBH β-ionone ring primarily by Gly^{181} → Ser replacement (as in VChR1,

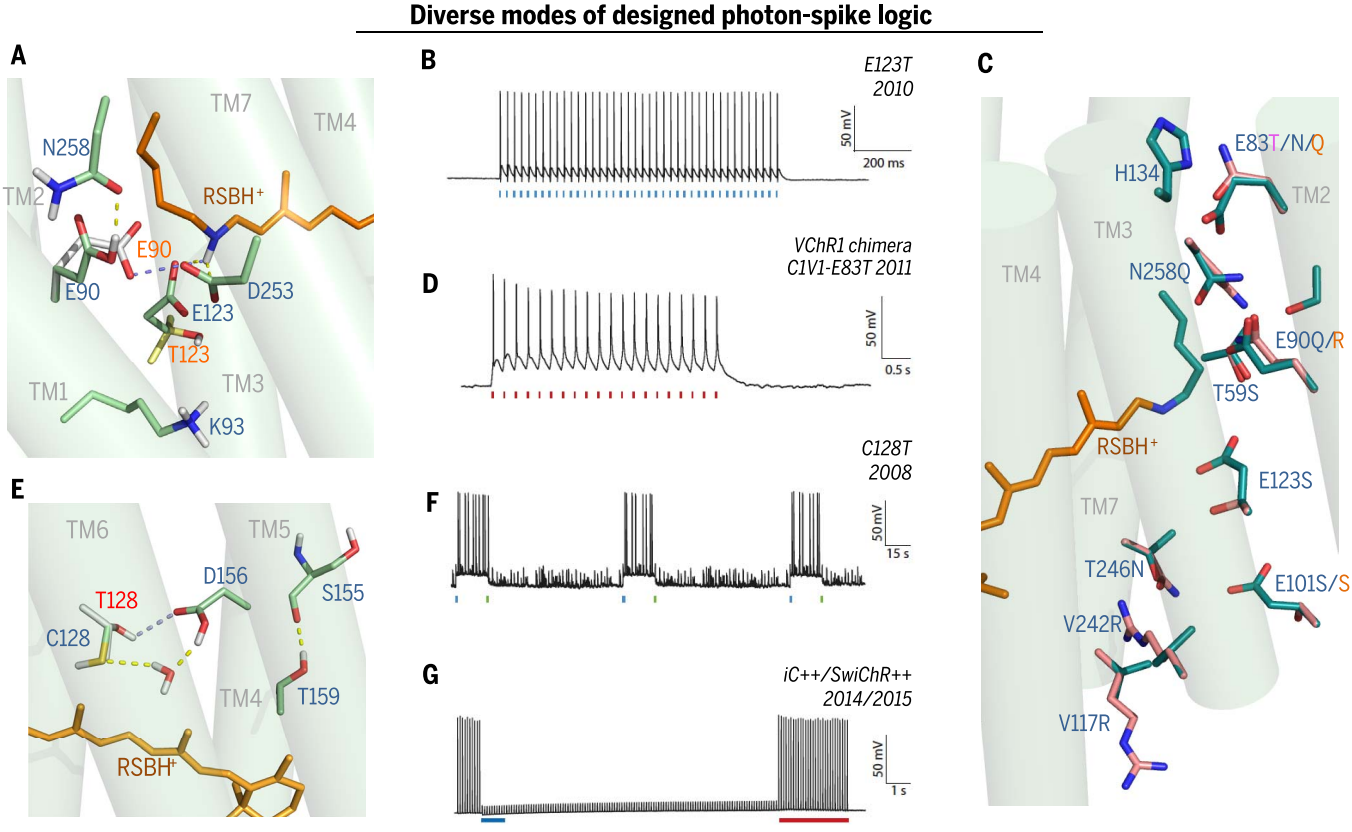


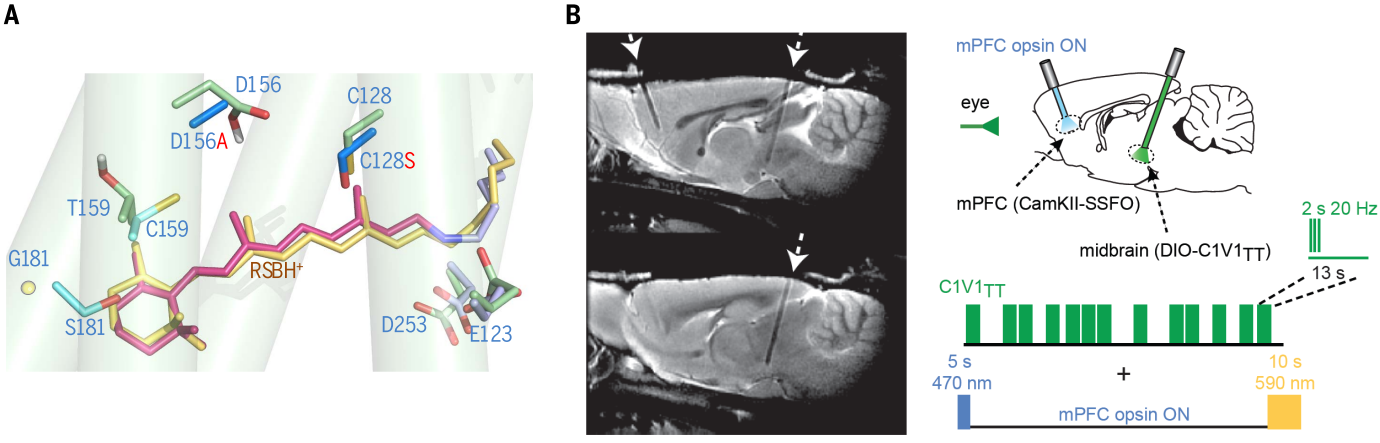
Fig. 4. Diverse modes of photon-spike transduction logic with underlying structural design. (**A**) Snapshot of ChR active site (MD calculation from C1C2 structure 3UG9, with predicted repositioning of side chains resulting from Glu^{123} → Thr (E123T) mutation in red lettering; E123T causes inward flipping of Glu^{90} to compensate for the lost Glu^{123} counterion (43) of the $RSBH^+$, additionally preventing inactivation. (**B**) Photon-spike transduction mode arising from ChETA mutation [ChR2-E123T variant (45)]: single blue flash–single spike coupling with high speed and high fidelity. Pore redesign implements faster closure after light-off, permitting rapid firing [e.g., 200-Hz trains in interneurons (45)]. (**C**) Pore residues (from C1C2 structure 3UG9) altered in spectral and selectivity variants. Modification of inner gate in red-activated C1V1-E83T variant (magenta letter–designated mutation; ChR2 numbering) (54). Selectivity variants are shown as original cation-conducting C1C2 pore residues and modifications to create the Cl^--selective iC++ [new pink side chains overlaid on original C1C2 green side chain positioning; blue letters denote iC++ mutations (62, 64)] or iChloC [orange letter–denoted mutations (93, 94)]. (**D**) Photon-spike transduction mode arising in *Volvox*-derived C1V1-E83T: single red flash–single spike coupling (54) with moderately high speed/fidelity; later *Volvox* derivative bReaChES exhibits faster responses with ChETA modification for accelerated channel closure (77) (not shown). (**E**) Snapshot of most likely structure of the DC-pair region in C1C2 (blue lettering) and the C128T (Cys^{128} → Thr) variant (red lettering) based on MD calculation (100) of restructured hydrogen-bonding network [yellow → blue dashed-line transition represents this SFO-mutation (52, 54) transition] and modified TM3-TM4 interaction (100), resulting in extension of open-state lifetime (52–54) and many-orders-of-magnitude-increased light sensitivity of expressing cells (42, 52, 54). (**F**) Photon-spike transduction mode arising from C128T (SFO) mutation is bistable, ultra–light-sensitive, two-color switchable, and excitatory [note blue light actuation and green light termination (52, 54)]. (**G**) Photon-spike transduction mode arising from adding the Cys^{128} SFO mutation (E) to Cl^--selective iC++ mutations (C) to create (64) SwiChR++ provides ultra–light sensitivity and is bistable, two-color switchable, and inhibitory under typical conditions (62, 64).

ReaChR, and Chrimson), and/or Thr[159] → Cys (or Met), Leu[162] → Cys, and Phe[178] → Tyr replacements (ChR2 numbering, Fig. 1), and/or by polarity reduction near the RSBH as in C1V1$_{TT}$ (Fig. 1 and Fig. 4, A and B). In the C1C2 crystal structure "frozen" snapshot, counterions Glu[123] and Asp[253] are directly connected to the RSBH+ proton without a water bridge, but molecular dynamics (MD) calculations reveal an active site with highly dynamic sampling of hydrogen-bonding patterns, with and without water bridges, separated only by tiny energetic differences (*81*). Red (lower-energy) photons suffice to induce deprotonation/isomerization in RBPs with high polarity near the β-ionone and low polarity at the RSBH [as positive charge shifts toward the ring (*69*, *70*); Figs. 4 and 5]. Although this principle has now been demonstrated, color tuning is still challenging, because high polarity near the β-ionone may reduce retinal-binding affinity, whereas low polarity near the RSBH lowers Schiff-base pK_a (favoring deprotonation), both of which could impair function. The ChR pore complex engages in long-range coupling interactions from the RBP to distant residues (Fig. 5); among other examples, substantial pH-dependent bathochromic color shifts were found to be associated with the presence of Glu[48] (using ChR2 numbering) at the N terminus of TM1 in redshifted ChR1, PsChR1, VChR1, DChR1, and relatives (Fig. 1). Precise mechanisms, although not known, could involve long-range coupling or tilting of the flexible TM1; causality is supported by observations that Glu[48] → Gln inhibited pH dependence of red shift, whereas Glu[48] → Ala (corresponding to blue-responsive ChR2; Fig. 1) instead stabilized the blue-absorbing form (*82*, *83*). Future advances in long-range molecular modeling methodology will build on these observations alongside current and emerging high-resolution structural and MD work.

The precise dynamical sequence of pore side chain repositioning will continue to serve as a subject of intense investigation and modeling. RBP amino acid replacements are expected to change energy profiles of both the electronic ground and excited states, with consequences for dynamics (influencing, besides spectrum and photocycle kinetics, the efficiency of retinal isomerization as with Glu[123] and Cys[128]). Several new structures, including additional red-shifted ChRs, are likely to emerge that will add finer-grained details to this intriguing landscape.

Selectivity variants

ChR ion selectivity is important in understanding the behavioral ecology of motile algae (*10*) and in achieving bidirectional impact on neuronal activity. Nonselective cation flux elicits excitation in neural systems via membrane depolarization, whereas anion or K^+ selectivity typically provides inhibition by clamping membrane potentials below the spike-firing threshold and reducing membrane resistance. *Chlamydomonas* ChRs naturally conduct cations, with modest inward rectification and cation species preference: $Li^+ > Na^+ > K^+ > Cs^+ >> Ca^{2+} > Mg^{2+}$ (*13*, *40*, *83*). Most ChRs are highly selective for protons, with $P_{H^+}/P_{Na^+} \sim 2 \times 10^6$ to 6×10^6 for ChR2 (*13*, *51*), comparable to that of highly selective mammalian proton channels (e.g., H^+H_v1; $P_{H^+}/P_{Na^+} \geq 10^6$) (*84*). For proton selectivity in algal ChRs, this level



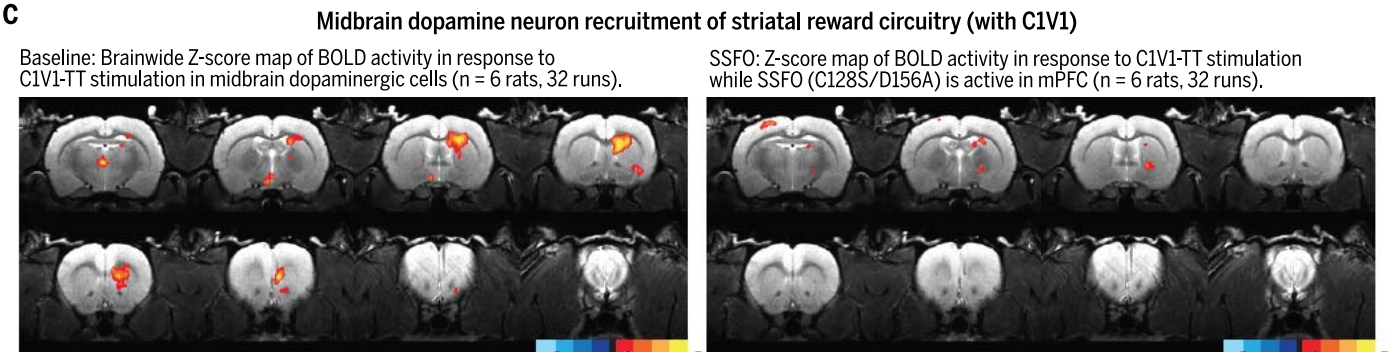


Fig. 5. Causal underpinnings of depression-related symptomatology identified via algal channel structure discovery and redesign. (**A**) Design of major opsin classes used together to identify brainwide dynamics of anhedonia (C1V1$_{TT}$/SSFO) (*56*): Model of red-shifted C1V1 RBP (red RSBH+; C1V1-specific side chains in cyan, based on 3UG9 structure) overlying modeled blue-responsive RBP (yellow RSBH+; green side chains, energy-optimized/calculated structure). Nearby DC-pair (SSFO) double mutant (D156A/C128S; new side chains in dark blue) shown (to illustrate relative positioning) that in the blue-responsive RBP confers stable activity shifts and blood oxygen level–dependent (BOLD) signal acquisition in MRI without heating or other artifacts. Expected further rearrangements and water influx are not shown. (**B**) Left: Tracks in rat brain for opsin injection and fiber-optic light access to mPFC (left arrow) and midbrain dopamine neurons (right arrow). Right: Testing causal influence of elevated mPFC activity (with blue-on/yellow-off SSFO) over communication within reward circuitry (from midbrain dopamine neurons controlled with C1V1). (**C**) Probing second-order brainwide dynamics. Natural prominent BOLD signal in dorsal and ventral striatum (left) recruited by dopamine neurons is potently suppressed (right) by mPFC excitability shift (*56*).

of preference suffices (85) because Na^+ becomes essentially irrelevant for freshwater algae in low-Na^+ environments. However, in mammalian brains with ~130 mM Na^+ at pH 7.5, Na^+ becomes a dominant component, modulated by voltage-dependent competition among protons/Na^+/K^+ and by weak Mg^{2+} block of proton permeation (40).

The C1C2 structure (15) alone was necessary but not sufficient for achieving insight into selective permeation, which required not only a crystal structure but also subsequent detailed structure-guided mechanistic studies. The magnitude of the mystery appeared even greater upon structure determination because few water molecules and no internally bound ions or high-affinity ion-binding sites were seen (15); moreover, the pore was large and disordered (relative to highly structured pores of Shaker and other K^+ channels) in the crystal (despite high-resolution refinement of the rigid RBP) (15), raising questions about the mechanism of cation conductance/selectivity. Because proton conductances can involve proton transfer reactions along residues forming a biological "proton wire," candidate protonatable pore-flanking residues were tested by replacement, with attention to five glutamates of helix 2 (E1 to E5, Fig. 1). Replacement of E3 [hydrogen-bonded to Ser^{63} and Asn^{258} forming the central gate (86, 87); ChR2 residue numbering] had the largest impact. Replacement of inner-gate His^{134}/E2 or access-channel E4/E5 also reduced H conductance (62, 64, 88, 89). Further support for this proton-wire hypothesis is derived from cryptophyte anion-conducting ChRs (90), which show reduced H^+ conductance with fewer helix-2 glutamates (90); interestingly, these glutamates have little influence on kinetics, and only mutation of E1 decelerates closing (88, 91).

ChR Ca^{2+} flux is presumed to be important for flagellar regulation, whereas Na^+/K^+ permeance could be incidental to more functionally relevant Ca^{2+}/H^+ flux in cation-conducting ChRs. Indeed, some ChRs (e.g., *Dunaliella salina*) are virtually H^+-selective (1). Incidental Na^+ flux might bring little energetic cost in freshwater algae (for pumping to restore small Na^+ gradients) and may arise as channels evolve within nonchannel gene families such as the pump-type microbial rhodopsins (perhaps after gene transfer in niches wherein archaea and algae are intimately linked, as with *Halobacteria salinarum* and *Dunaliella salina* in high-salt environments). The versatile ChR pore without high-affinity ion-binding sites (15) contrasts with naturally occurring K^+- or Ca^{2+}-channel selectivity mechanisms, which use tetrameric and symmetric structures to achieve coordinated ion-binding sites that mimic hydration shells or direct symmetric chelation of ions, respectively.

How, then, do ChRs achieve cation selectivity? The C1C2 crystal structure (15) revealed a preponderance of intracellular vestibule basic residues (64, 92), but attempts to modify conduction pathway access here and elsewhere (although successful in quantitatively shifting ion permeation ratios) did not alter cation selectivity (13). However, the predicted electronegative environment of the entire conduction pathway in cation-selective C1C2 (64, 92) suggested a pore-lining surface electrostatic model and new possibilities for testing mechanisms of selective permeation. In 2014, a direct test (remodeling the C1C2 pore for surface electropositivity) confirmed this new model (62), resulting in a Cl^--selective ChR (iC1C2) that was inhibitory in neural systems (62). At the same time, different mutations (including negative-to-positive replacement of the central gate Glu^{90} with Lys^+ or Arg^+) were used to create a Cl^--selective ChR2 (93), with the unifying theme of similar-direction effects on predicted internal pore electrostatics (93). Further mutagenesis of both variants [guided by the new model, in which complete elimination of proton conductance also required mutation of one or two putative proton-wire pore glutamates (64, 94)] resulted in highly Cl^--selective ChRs [iChloC and iC++; Fig. 4G (64, 94)] now widely used for studies of animal behavior (conferring optogenetic inhibition in typical situations wherein internal Cl^- concentration is low) (64, 95, 96).

Naturally occurring Cl^--selective ChRs (GtACR1 and GtACR2) were described from cryptophyte algae the year after the first designed Cl^--selective ChR was reported; these were termed anion channelrhodopsins (ACRs) (90). Even though ACRs are evolutionarily remote from cation-conducting ChRs with more amino acid replacements than the designed Cl^--conducting ChRs, these new ChRs independently fit the earlier pore model as well, with the unifying principle across all Cl^--selective ChRs corresponding to a net effect on pore-lining electrostatics (92). The universality of the selectivity mechanism is underscored by the fact that no single charge seems crucial for Cl^- selectivity, as long as net pore electrostatics are preserved (92). ChR Glu^{90} is invariant in all naturally occurring ChRs (Glu^{64} in GtACR2 numbering), but this negative charge is compensated in anion-conducting GtACR2 via numerous charge-altering replacements of other pore-facing glutamate residues found in cation-conducting ChRs (changed in GtACR2 to non-charged residues: Ser^{57}, Thr^{67}, Ala^{71}, Asn^{75}, and Ser^{93}) (92). Moreover, Glu^{90} is not essential for Cl^- flux; both families of engineered Cl^--selective ChRs (iC1C2, ChloC) lack glutamate at this position (62, 93). Finally, even altering this glutamate in naturally occurring GtACR1 ($Glu^{68} \rightarrow$ Gln) has no detectable effect on photocurrent magnitude or kinetics under physiological conditions (91). Although the ACR primary sequence suggests rearrangement at both gates, an E3 at the central gate is retained (Fig. 1). The comparative value of natural variants is substantial, but structural and spectroscopic data remain to be acquired.

The intense interest that led to resolving this fundamental question of selectivity had the additional benefit of driving the creation of new optogenetic tools, enabling the first round of studies using Cl^--selective ChRs for inhibitory optogenetics in animal behavior (64, 95, 96). Moreover, integration of diverse structural insights has begun; the SFO kinetic principle was discovered to be portable to inhibitory ChRs, creating bistable and extremely light-sensitive inhibitory ChRs (SwiChR and SwiChR++) (62, 64) that have found utility in studies of mouse behavior (60).

Outlook

The ChR light-gated pore is unique among biological structures. The investigation, modeling, and redesign of atomic-scale structure-function relationships governing this light-to-current converter led not only to advances in neuroscience but also to sophisticated understanding of the underlying chemical processes. These light-gated channels will continue to fascinate (and occupy a privileged position within ion-channel research) because only light-gated systems allow structure-function analyses over time scales of femtoseconds (10^{-15} s) to seconds or more (97).

The role of ChR in neuroscience may continue to be special as well, building on precision redesign for new roles (Fig. 4). Diverse modes of photon-spike logic are now accessible (four examples in Fig. 4, each alongside key pore-modeling and pore-redesign features required to create the new logic). Even psychiatry has yielded some of its deepest mysteries to ChR pore structural insights (Fig. 5), as exemplified by studies of circuit dynamics underlying the core depression symptom of anhedonia (59) in which natural reward responses central to the behavior of all animals are lost. The presence of anhedonia allows a diagnosis of major depressive disorder even without depressed mood, but causal circuit dynamics–level understanding had remained elusive.

The experiments summarized in Fig. 5 used a combination of designed opsins arising from structure-function ChR discoveries (56). The blue light–activated SSFO (DC pair–modified for bistable behavior on a 30-min time scale) allowed causal analysis of brainwide dynamics in awake rats during fMRI scanning (56). SSFO modulation in mPFC was combined with modulation of neural elements across the brain using $C1V1_{TT}$ [containing no ChR2 sequence but with chimeric elements of VChR1/ChR1, RBP point mutations for redshifted compatibility (54) with SSFO, and the ChETA (45) mutation] (56). Figure 5A shows elements of the pore redesign that enabled this approach; Fig. 5B shows the strategy for brain-spanning interrogation of reward circuitry. Elevated mPFC activity (to mimic a pattern seen in depression with anhedonia) altered the ability of midbrain dopamine neurons to recruit striatal reward circuitry (Fig. 5C)—a higher-order brainwide form of modulation by which the evolutionarily advanced frontal neocortex is capable of modifying interactions between two distant subcortical structures—thereby controlling reward-mediating physiology and behavior (56).

In addition to these and other insights into adaptive and maladaptive behavior, basic insights into ion channel biophysics and evolution have emerged. Since the resolution of the crystal structure of ChR and the demonstration of the pore surface electrostatic mechanism for Cl^- selectivity, two recent cryo-EM structures subsequently emerged for unrelated Cl^- channels that (like ChR) have arisen within larger nonchannel protein families (98, 99). The cystic fibrosis transmembrane

conductance regulator (CFTR) is a Cl^- channel within the nonchannel ABC transporter family. Cryo-EM (*98*) revealed a putative Cl^- conduction pathway with striking parallels to the unrelated 7TM Cl^--conducting ChRs with its wide pore, lack of high-affinity identified Cl^- binding sites (as detectable in lower-resolution cryo-EM), and largely continuous positive surface electrostatics along the conduction pathway. Similarly, CLC-K is a Cl^- channel within a broad family of 12TM CLC proteins that includes nonchannel Cl^-/H^+ exchangers. CLC-K, like CFTR, is distinct from ChRs in topology (12-TM protein dimer). Cryo-EM (*99*) for CLC-K revealed a pattern of positively charged arginines in the conduction pathway (with no high-affinity binding sites detectable in cryo-EM). Together with the earlier ChR discoveries, a model emerges by which Cl^--conducting channels can evolve using low-affinity surface electrostatic pathways, within nonchannel families that may not otherwise be well suited (by symmetry or other properties) to give rise to highly selective channels.

Much more remains to be uncovered regarding the operation of these unparalleled biological machines, including mechanisms of long-range molecular tuning of color, kinetics, and selectivity. Understanding will continue to be refined with new structural information and ongoing advances in modeling of molecular dynamics, which will also likely continue to drive neuroscience. For light-activated ChR pores, studying the interplay of form and function has illuminated the governing principles of the channel, the basic behavioral ecology of algae, the evolution of ion channels, and the fundamental circuit underpinnings of animal behavior in health and disease—remarkable achievements for a light-activated molecule from a single-celled plant.

REFERENCES AND NOTES

1. F. Zhang *et al*., The microbial opsin family of optogenetic tools. *Cell* **147**, 1446–1457 (2011). doi: 10.1016/j.cell.2011.12.004; pmid: 22196724
2. O. P. Ernst *et al*., Microbial and animal rhodopsins: Structures, functions, and molecular mechanisms. *Chem. Rev.* **114**, 126–163 (2014). doi: 10.1021/cr4003769; pmid: 24364740
3. C. K. Kim, A. Adhikari, K. Deisseroth, Integration of optogenetics with complementary methodologies in systems neuroscience. *Nat. Rev. Neurosci.* **18**, 222–235 (2017). doi: 10.1038/nrn.2017.15; pmid: 28303019
4. K. W. Foster, R. D. Smyth, Light antennas in phototactic algae. *Microbiol. Rev.* **44**, 572–630 (1980). pmid: 7010112
5. A. Faminzin, Die Wirkung des Lichtes auf die Bewegung der *Chlamidomonas pulvisculus* Ehr., *Euglena viridis* Ehr. und *Oscillatoria insignis* Tw. In *Melanges Biologiques tires du Bulletin de l'Ácademie Imperial des Sciences De St.-Petersbourg* (1866), pp. 73–93.
6. F. F. Litvin, O. A. Sineshchekov, V. A. Sineshchekov, Photoreceptor electric potential in the phototaxis of the alga *Haematococcus pluvialis*. *Nature* **271**, 476–478 (1978). doi: 10.1038/271476a0; pmid: 628427
7. H. Harz, P. Hegemann, Rhodopsin-regulated calcium currents in *Chlamydomonas*. *Nature* **351**, 489–491 (1991). doi: 10.1038/351489a0
8. D. Oesterhelt, W. Stoeckenius, Rhodopsin-like protein from the purple membrane of *Halobacterium halobium*. *Nat. New Biol.* **233**, 149–152 (1971). doi: 10.1038/newbio233149a0; pmid: 4940442
9. T. R. Herrmann, G. W. Rayfield, A measurement of the proton pump current generated by bacteriorhodopsin in black lipid membranes. *Biochim. Biophys. Acta* **443**, 623–628 (1976). doi: 10.1016/0005-2736(76)90482-X; pmid: 963073
10. K. W. Foster *et al*., A rhodopsin is the functional photoreceptor for phototaxis in the unicellular eukaryote *Chlamydomonas*. *Nature* **311**, 756–759 (1984). doi: 10.1038/311756a0; pmid: 6493336
11. F. J. Braun, P. Hegemann, Direct measurement of cytosolic calcium and pH in living *Chlamydomonas reinhardtii* cells. *Eur. J. Cell Biol.* **78**, 199–208 (1999). doi: 10.1016/S0171-9335(99)80099-5; pmid: 10219570
12. G. Nagel *et al*., Channelrhodopsin-1: A light-gated proton channel in green algae. *Science* **296**, 2395–2398 (2002). doi: 10.1126/science.1072068; pmid: 12089443
13. G. Nagel *et al*., Channelrhodopsin-2, a directly light-gated cation-selective membrane channel. *Proc. Natl. Acad. Sci. U.S.A.* **100**, 13940–13945 (2003). doi: 10.1073/pnas.1936192100; pmid: 14615590
14. K. Deisseroth, Optogenetics: 10 years of microbial opsins in neuroscience. *Nat. Neurosci.* **18**, 1213–1225 (2015). doi: 10.1038/nn.4091; pmid: 26308982
15. H. E. Kato *et al*., Crystal structure of the channelrhodopsin light-gated cation channel. *Nature* **482**, 369–374 (2012). doi: 10.1038/nature10870; pmid: 22266941
16. M. Iseki *et al*., A blue-light-activated adenylyl cyclase mediates photoavoidance in *Euglena gracilis*. *Nature* **415**, 1047–1051 (2002). doi: 10.1038/4151047a; pmid: 11875575
17. B. D. Engel *et al*., Native architecture of the *Chlamydomonas* chloroplast revealed by in situ cryo-electron tomography. *eLife* **4**, e04889 (2015). doi: 10.7554/eLife.04889
18. P. Berthold *et al*., Channelrhodopsin-1 initiates phototaxis and photophobic responses in *Chlamydomonas* by immediate light-induced depolarization. *Plant Cell* **20**, 1665–1677 (2008). doi: 10.1105/tpc.108.057919; pmid: 18552201
19. T. M. Mittelmeier, J. S. Boyd, M. R. Lamb, C. L. Dieckmann, Asymmetric properties of the *Chlamydomonas reinhardtii* cytoskeleton direct rhodopsin photoreceptor localization. *J.Cell Biol.* **193**, 741–753 (2011). doi: 10.1083/jcb.201009131; pmid: 21555459
20. K. Y. Wan, K. C. Leptos, R. E. Goldstein, Lag, lock, sync, slip: The many 'phases' of coupled flagella. *J. R. Soc. Interface* **11**, 20131160 (2014). doi: 10.1098/rsif.2013.1160; pmid: 24573332
21. F. Crick, The impact of molecular biology on neuroscience. *Philos. Trans. R. Soc. B* **354**, 2021–2025 (1999). doi: 10.1098/rstb.1999.0541; pmid: 10670022
22. I. C. Farber, A. Grinvald, Identification of presynaptic neurons by laser photostimulation. *Science* **222**, 1025–1027 (1983). doi: 10.1126/science.6648515; pmid: 6648515
23. R. L. Fork, Laser stimulation of nerve cells in *Aplysia*. *Science* **171**, 907–908 (1971). doi: 10.1126/science.171.3974.907; pmid: 5541653
24. B. V. Zemelman, G. A. Lee, M. Ng, G. Miesenböck, Selective photostimulation of genetically chARGed neurons. *Neuron* **33**, 15–22 (2002). doi: 10.1016/S0896-6273(01)00574-8; pmid: 11779476
25. M. Banghart, K. Borges, E. Isacoff, D. Trauner, R. H. Kramer, Light-activated ion channels for remote control of neuronal firing. *Nat. Neurosci.* **7**, 1381–1386 (2004). doi: 10.1038/nn1356; pmid: 15558062
26. S. Q. Lima, G. Miesenböck, Remote control of behavior through genetically targeted photostimulation of neurons. *Cell* **121**, 141–152 (2005). doi: 10.1016/j.cell.2005.02.004; pmid: 15820685
27. E. S. Boyden, F. Zhang, E. Bamberg, G. Nagel, K. Deisseroth, Millisecond-timescale, genetically targeted optical control of neural activity. *Nat. Neurosci.* **8**, 1263–1268 (2005). doi: 10.1038/nn1525; pmid: 16116447
28. X. Li *et al*., Fast noninvasive activation and inhibition of neural and network activity by vertebrate rhodopsin and green algae channelrhodopsin. *Proc. Natl. Acad. Sci. U.S.A.* **102**, 17816–17821 (2005). doi: 10.1073/pnas.0509030102; pmid: 16306259
29. T. Ishizuka, M. Kakuda, R. Araki, H. Yawo, Kinetic evaluation of photosensitivity in genetically engineered neurons expressing green algae light-gated channels. *Neurosci. Res.* **54**, 85–94 (2006). doi: 10.1016/j.neures.2005.10.009; pmid: 16298005
30. G. Nagel *et al*., Light activation of channelrhodopsin-2 in excitable cells of *Caenorhabditis elegans* triggers rapid behavioral responses. *Curr. Biol.* **15**, 2279–2284 (2005). doi: 10.1016/j.cub.2005.11.032; pmid: 16360690
31. A. Bi *et al*., Ectopic expression of a microbial-type rhodopsin restores visual responses in mice with photoreceptor degeneration. *Neuron* **50**, 23–33 (2006). doi: 10.1016/j.neuron.2006.02.026; pmid: 16600853
32. A. M. Aravanis *et al*., An optical neural interface: In vivo control of rodent motor cortex with integrated fiberoptic and optogenetic technology. *J. Neural Eng.* **4**, S143–S156 (2007). doi: 10.1088/1741-2560/4/3/S02; pmid: 17873414
33. A. R. Adamantidis, F. Zhang, A. M. Aravanis, K. Deisseroth, L. de Lecea, Neural substrates of awakening probed with optogenetic control of hypocretin neurons. *Nature* **450**, 420–424 (2007). doi: 10.1038/nature06310; pmid: 17943086
34. V. Gradinaru, M. Mogri, K. R. Thompson, J. M. Henderson, K. Deisseroth, Optical deconstruction of parkinsonian neural circuitry. *Science* **324**, 354–359 (2009). doi: 10.1126/science.1167093; pmid: 19299587
35. H. C. Tsai *et al*., Phasic firing in dopaminergic neurons is sufficient for behavioral conditioning. *Science* **324**, 1080–1084 (2009). doi: 10.1126/science.1168878; pmid: 19389999
36. M. Müller, C. Bamann, E. Bamberg, W. Kühlbrandt, Projection structure of channelrhodopsin-2 at 6 Å resolution by electron crystallography. *J. Mol. Biol.* **414**, 86–95 (2011). doi: 10.1016/j.jmb.2011.09.049; pmid: 22001017
37. S. Bruun *et al*., Light-dark adaptation of channelrhodopsin involves photoconversion between the all-*trans* and 13-*cis* retinal isomers. *Biochemistry* **54**, 5389–5400 (2015). doi: 10.1021/acs.biochem.5b00597; pmid: 26237332
38. R. Vogel *et al*., Deactivation of rhodopsin in the transition from the signaling state meta II to meta III involves a thermal isomerization of the retinal chromophore C=D. *Biochemistry* **42**, 9863–9874 (2003). doi: 10.1021/bi034684+; pmid: 12924935
39. G. S. Harbison *et al*., Dark-adapted bacteriorhodopsin contains 13-cis, 15-syn and all-trans, 15-anti retinal Schiff bases. *Proc. Natl. Acad. Sci. U.S.A.* **81**, 1706–1709 (1984). doi: 10.1073/pnas.81.6.1706; pmid: 6584904
40. F. Schneider, D. Gradmann, P. Hegemann, Ion selectivity and competition in channelrhodopsins. *Biophys. J.* **105**, 91–100 (2013). doi: 10.1016/j.bpj.2013.05.042; pmid: 23823227
41. F. Schneider, C. Grimm, P. Hegemann, Biophysics of channelrhodopsin. *Annu. Rev. Biophys.* **44**, 167–186 (2015). doi: 10.1146/annurev-biophys-060414-034014; pmid: 26098512
42. J. Mattis *et al*., Principles for applying optogenetic tools derived from direct comparative analysis of microbial opsins. *Nat. Methods* **9**, 159–172 (2011). doi: 10.1038/nmeth.1808; pmid: 22179551
43. J. Kuhne *et al*., Early formation of the ion-conducting pore in channelrhodopsin-2. *Angew. Chem. Int. Ed.* **54**, 4953–4957 (2015). doi: 10.1002/anie.201410180; pmid: 25537168
44. V. A. Lórenz-Fonfría *et al*., Temporal evolution of helix hydration in a light-gated ion channel correlates with ion conductance. *Proc. Natl. Acad. Sci. U.S.A.* **112**, E5796–E5804 (2015). doi: 10.1073/pnas.1511462112; pmid: 26460012
45. L. A. Gunaydin *et al*., Ultrafast optogenetic control. *Nat. Neurosci.* **13**, 387–392 (2010). doi: 10.1038/nn.2495; pmid: 20081849
46. F. Assaf, Y. Schiller, The antiepileptic and ictogenic effects of optogenetic neurostimulation of PV-expressing interneurons. *J. Neurophysiol.* **116**, 1694–1704 (2016). doi: 10.1152/jn.00744.2015; pmid: 27486107
47. S. Jego *et al*., Optogenetic identification of a rapid eye movement sleep modulatory circuit in the hypothalamus. *Nat. Neurosci.* **16**, 1637–1643 (2013). doi: 10.1038/nn.3522; pmid: 24056699
48. O. Klavir, M. Prigge, A. Sarel, R. Paz, O. Yizhar, Manipulating fear associations via optogenetic modulation of amygdala inputs to prefrontal cortex. *Nat. Neurosci.* **20**, 836–844 (2017). doi: 10.1038/nn.4523; pmid: 28288126
49. M. Carus-Cadavieco *et al*., Gamma oscillations organize top-down signalling to hypothalamus and enable food seeking. *Nature* **542**, 232–236 (2017). doi: 10.1038/nature21066; pmid: 28146472
50. L. Grosenick, J. H. Marshel, K. Deisseroth, Closed-loop and activity-guided optogenetic control. *Neuron* **86**, 106–139 (2015). doi: 10.1016/j.neuron.2015.03.034; pmid: 25856490
51. A. Berndt *et al*., High-efficiency channelrhodopsins for fast neuronal stimulation at low light levels. *Proc. Natl. Acad. Sci. U.S.A.* **108**, 7595–7600 (2011). doi: 10.1073/pnas.1017210108; pmid: 21504945
52. A. Berndt, O. Yizhar, L. A. Gunaydin, P. Hegemann, K. Deisseroth, Bi-stable neural state switches. *Nat. Neurosci.* **12**, 229–234 (2009). doi: 10.1038/nn.2247; pmid: 19079251

53. C. Bamann, R. Gueta, S. Kleinlogel, G. Nagel, E. Bamberg, Structural guidance of the photocycle of channelrhodopsin-2 by an interhelical hydrogen bond. *Biochemistry* **49**, 267–278 (2010). doi: 10.1021/bi901634p; pmid: 20000562
54. O. Yizhar *et al*., Neocortical excitation/inhibition balance in information processing and social dysfunction. *Nature* **477**, 171–178 (2011). doi: 10.1038/nature10360; pmid: 21796121
55. L. I. Schmitt *et al*., Thalamic amplification of cortical connectivity sustains attentional control. *Nature* **545**, 219–223 (2017). doi: 10.1038/nature22073; pmid: 28467827
56. E. A. Ferenczi *et al*., Prefrontal cortical regulation of brainwide circuit dynamics and reward-related behavior. *Science* **351**, aac9698 (2016). doi: 10.1126/science.aac9698; pmid: 26722001
57. C. L. Tan *et al*., Warm-sensitive neurons that control body temperature. *Cell* **167**, 47–59.e15 (2016). doi: 10.1016/j.cell.2016.08.028; pmid: 27616062
58. J. M. Sorokin *et al*., Bidirectional control of generalized epilepsy networks via rapid real-time switching of firing mode. *Neuron* **93**, 194–210 (2017). doi: 10.1016/j.neuron.2016.11.026; pmid: 27989462
59. H. Makino, T. Komiyama, Learning enhances the relative impact of top-down processing in the visual cortex. *Nat. Neurosci.* **18**, 1116–1122 (2015). doi: 10.1038/nn.4061; pmid: 26167904
60. S. M. Iyer *et al*., Optogenetic and chemogenetic strategies for sustained inhibition of pain. *Sci. Rep.* **6**, 30570 (2016). doi: 10.1038/srep30570; pmid: 27484850
61. V. Haikala, M. Joesch, A. Borst, A. S. Mauss, Optogenetic control of fly optomotor responses. *J. Neurosci.* **33**, 13927–13934 (2013). doi: 10.1523/JNEUROSCI.0340-13.2013; pmid: 23966712
62. A. Berndt, S. Y. Lee, C. Ramakrishnan, K. Deisseroth, Structure-guided transformation of channelrhodopsin into a light-activated chloride channel. *Science* **344**, 420–424 (2014). doi: 10.1126/science.1252367; pmid: 24763591
63. R. Prakash *et al*., Two-photon optogenetic toolbox for fast inhibition, excitation and bistable modulation. *Nat. Methods* **9**, 1171–1179 (2012). doi: 10.1038/nmeth.2215; pmid: 23169303
64. A. Berndt *et al*., Structural foundations of optogenetics: Determinants of channelrhodopsin ion selectivity. *Proc. Natl. Acad. Sci. U.S.A.* **113**, 822–829 (2016). doi: 10.1073/pnas.1523341113; pmid: 26699459
65. N. C. Klapoetke *et al*., Independent optical excitation of distinct neural populations. *Nat. Methods* **11**, 338–346 (2014). doi: 10.1038/nmeth.2836; pmid: 24509633
66. J. Marshall, T. W. Cronin, S. Kleinlogel, Stomatopod eye structure and function: A review. *Arthropod Struct. Dev.* **36**, 420–448 (2007). doi: 10.1016/j.asd.2007.01.006; pmid: 18089120
67. W. Wang *et al*., Tuning the electronic absorption of protein-embedded all-trans-retinal. *Science* **338**, 1340–1343 (2012). doi: 10.1126/science.1226135; pmid: 23224553
68. H. E. Kato *et al*., Atomistic design of microbial opsin-based blue-shifted optogenetics tools. *Nat. Commun.* **6**, 7177 (2015). doi: 10.1038/ncomms8177; pmid: 25975962
69. E. Kloppmann, T. Becker, G. M. Ullmann, Electrostatic potential at the retinal of three archaeal rhodopsins: Implications for their different absorption spectra. *Proteins* **61**, 953–965 (2005). doi: 10.1002/prot.20744; pmid: 16247786
70. F. Zhang *et al*., Red-shifted optogenetic excitation: A tool for fast neural control derived from *Volvox carteri*. *Nat. Neurosci.* **11**, 631–633 (2008). doi: 10.1038/nn.2120; pmid: 18432196
71. V. Buss, Inherent chirality of the retinal chromophore in rhodopsin—A nonempirical theoretical analysis of chiroptical data. *Chirality* **13**, 13–23 (2001). doi: 10.1002/1520-636X(2001)13:1<13::AID-CHIR4>3.0.CO;2-3; pmid: 11135409
72. V. Gradinaru *et al*., Molecular and cellular approaches for diversifying and extending optogenetics. *Cell* **141**, 154–165 (2010). doi: 10.1016/j.cell.2010.02.037; pmid: 20303157
73. H. Wang *et al*., Molecular determinants differentiating photocurrent properties of two channelrhodopsins from chlamydomonas. *J. Biol. Chem.* **284**, 5685–5696 (2009). doi: 10.1074/jbc.M807632200; pmid: 19103605
74. J. Y. Lin, M. Z. Lin, P. Steinbach, R. Y. Tsien, Characterization of engineered channelrhodopsin variants with improved properties and kinetics. *Biophys. J.* **96**, 1803–1814 (2009). doi: 10.1016/j.bpj.2008.11.034; pmid: 19254539
75. J. P. Rickgauer, K. Deisseroth, D. W. Tank, Simultaneous cellular-resolution optical perturbation and imaging of place cell firing fields. *Nat. Neurosci.* **17**, 1816–1824 (2014). doi: 10.1038/nn.3866; pmid: 25402854
76. C. K. Kim *et al*., Simultaneous fast measurement of circuit dynamics at multiple sites across the mammalian brain. *Nat. Methods* **13**, 325–328 (2016). doi: 10.1038/nmeth.3770; pmid: 26878381
77. P. Rajasethupathy *et al*., Projections from neocortex mediate top-down control of memory retrieval. *Nature* **526**, 653–659 (2015). doi: 10.1038/nature15389; pmid: 26436451
78. L. Carrillo-Reid, W. Yang, Y. Bando, D. S. Peterka, R. Yuste, Imprinting and recalling cortical ensembles. *Science* **353**, 691–694 (2016). doi: 10.1126/science.aaf7560; pmid: 27516599
79. J. Y. Lin, P. M. Knutsen, A. Muller, D. Kleinfeld, R. Y. Tsien, ReaChR: A red-shifted variant of channelrhodopsin enables deep transcranial optogenetic excitation. *Nat. Neurosci.* **16**, 1499–1508 (2013). doi: 10.1038/nn.3502; pmid: 23995068
80. E. G. Govorunova, E. N. Spudich, C. E. Lane, O. A. Sineshchekov, J. L. Spudich, New channelrhodopsin with a red-shifted spectrum and rapid kinetics from *Mesostigma viride*. *MBio* **2**, e00115–e11 (2011). doi: 10.1128/mBio.00115-11; pmid: 21693637
81. Y. Guo *et al*., Active site structure and absorption spectrum of channelrhodopsin-2 wild-type and C128T mutant. *Chem. Sci.* **7**, 3879–3891 (2016). doi: 10.1039/C6SC00468G
82. S. P. Tsunoda, P. Hegemann, Glu 87 of channelrhodopsin-1 causes pH-dependent color tuning and fast photocurrent inactivation. *Photochem. Photobiol.* **85**, 564–569 (2009). doi: 10.1111/j.1751-1097.2008.00519.x; pmid: 19192197
83. E. G. Govorunova, O. A. Sineshchekov, H. Li, R. Janz, J. L. Spudich, Characterization of a highly efficient blue-shifted channelrhodopsin from the marine alga *Platymonas subcordiformis*. *J. Biol. Chem.* **288**, 29911–29922 (2013). doi: 10.1074/jbc.M113.505495; pmid: 23995841
84. N. Demaurex *et al*., Proton currents in human granulocytes: Regulation by membrane potential and intracellular pH. *J. Physiol.* **466**, 329–344 (1993). pmid: 7692041
85. S. Ehlenbeck, D. Gradmann, F.-J. Braun, P. Hegemann, Evidence for a light-induced H^+ conductance in the eye of the green alga *Chlamydomonas reinhardtii*. *Biophys. J.* **82**, 740–751 (2002). doi: 10.1016/S0006-3495(02)75436-2; pmid: 11806916
86. K. Eisenhauer *et al*., In channelrhodopsin-2 Glu-90 is crucial for ion selectivity and is deprotonated during the photocycle. *J. Biol. Chem.* **287**, 6904–6911 (2012). doi: 10.1074/jbc.M111.327700
87. K. Ruffert *et al*., Glutamate residue 90 in the predicted transmembrane domain 2 is crucial for cation flux through channelrhodopsin 2. *Biochem. Biophys. Res. Commun.* **410**, 737–743 (2011). pmid: 21683688
88. A. P. Plazzo *et al*., Bioinformatic and mutational analysis of channelrhodopsin-2 protein cation-conducting pathway. *J. Biol. Chem.* **287**, 4818–4825 (2012). doi: 10.1074/jbc.M111.326207; pmid: 22139833
89. J. Wietek, M. Broser, B. S. Krause, P. Hegemann, Identification of a natural green light absorbing chloride conducting channelrhodopsin from *Proteomonas sulcata*. *J. Biol. Chem.* **291**, 4121–4127 (2016). doi: 10.1074/jbc.M115.699637; pmid: 26740624
90. E. G. Govorunova, O. A. Sineshchekov, R. Janz, X. Liu, J. L. Spudich, Natural light-gated anion channels: A family of microbial rhodopsins for advanced optogenetics. *Science* **349**, 647–650 (2015). doi: 10.1126/science.aaa7484; pmid: 26113638
91. E. G. Govorunova, S. R. Cunha, O. A. Sineshchekov, J. L. Spudich, Anion channelrhodopsins for inhibitory cardiac optogenetics. *Sci. Rep.* **6**, 33530 (2016). doi: 10.1038/srep33530; pmid: 27628215
92. A. Berndt, K. Deisseroth, Expanding the optogenetics toolkit. *Science* **349**, 590–591 (2015). doi: 10.1126/science.aac7889; pmid: 26250674
93. J. Wietek *et al*., Conversion of channelrhodopsin into a light-gated chloride channel. *Science* **344**, 409–412 (2014). doi: 10.1126/science.1249375; pmid: 24674867
94. J. Wietek *et al*., An improved chloride-conducting channelrhodopsin for light-induced inhibition of neuronal activity in vivo. *Sci. Rep.* **5**, 14807 (2015). doi: 10.1038/srep14807; pmid: 26443033
95. H. Kim, S. Ährlund-Richter, X. Wang, K. Deisseroth, M. Carlén, Prefrontal parvalbumin neurons in control of attention. *Cell* **164**, 208–218 (2016). doi: 10.1016/j.cell.2015.11.038; pmid: 26771492
96. N. Takahashi, T. G. Oertner, P. Hegemann, M. E. Larkum, Active cortical dendrites modulate perception. *Science* **354**, 1587–1590 (2016). doi: 10.1126/science.aah6066; pmid: 28008068
97. E. Nango *et al*., A three-dimensional movie of structural changes in bacteriorhodopsin. *Science* **354**, 1552–1557 (2016). doi: 10.1126/science.aah3497; pmid: 28008064
98. F. Liu, Z. Zhang, L. Csanády, D. C. Gadsby, J. Chen, Molecular structure of the human CFTR ion channel. *Cell* **169**, 85–95.e8 (2017). doi: 10.1016/j.cell.2017.02.024; pmid: 28340353
99. E. Park, E. B. Campbell, R. MacKinnon, Structure of a CLC chloride ion channel by cryo-electron microscopy. *Nature* **541**, 500–505 (2017). doi: 10.1038/nature20812; pmid: 28002411
100. H. C. Watanabe, K. Welke, D. J. Sindhikara, P. Hegemann, M. Elstner, Towards an understanding of channelrhodopsin function: Simulations lead to novel insights of the channel mechanism. *J. Mol. Biol.* **425**, 1795–1814 (2013). doi: 10.1016/j.jmb.2013.01.033; pmid: 23376098

ACKNOWLEDGMENTS

We thank our co-workers for contributions to the analysis of ChR structure and function over the years, and for the interdisciplinary exchange of knowledge among the communities of algal biology, membrane biophysics, structural biology, neuroscience, and medicine. We also thank E. A. Lysenko (Institute of Plant Physiology, RAS Moscow) for the photograph of Faminzin; F. Beyle for the ChR2 structure homology file; A. Berndt, E. Ferenczi, H. Kato, Y. Kim, F. Schneider, J. Wietek, and O. Yizhar for comments; and S. Kelterborn for Fig. 2B. Supported by the Deutsche Forschungsgemeinschaft, European Research Council, and Hertie Foundation (P.H.) and by NIH, NSF, DARPA, the NOMIS Foundation, and the Else Kroner Fresenius Foundation (K.D.).

10.1126/science.aan5544

The form and function of channelrhodopsin

Karl Deisseroth and Peter Hegemann

Science **357** (6356), eaan5544.
DOI: 10.1126/science.aan5544

From biophysics to neuroscience tools

The channelrhodopsins and their distinctive light-activated ion channels have emerged as major tools in modern biological research. Deisseroth and Hegemann review the structural and functional properties of these protein photoreceptors. Mutagenesis and modeling studies, coupled with the reintroduction of modified channels into living systems, offer a profound understanding of how these channels work. The insights into the underlying basic science provide foundations for developing further applications in biology and medicine.

Science, this issue p. eaan5544